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Two new compounds from *Carthamus tinctorius*

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From the dried petals of *Carthamus tinctorius*, a new flavonoid, (2*R*)-4',5-dihydroxyl-6,7-di-*O*-β-D-glucopyranosyl flavanone (**1**) and a new aromatic glucoside, methyl-3-(4-*O*-β-D-glucopyranosylphenyl) propionate (**4**) were isolated along with four known compounds (2*S*)-4',5-dihydroxyl-6,7-di-*O*-β-D-glucopyranosyl flavanone (**2**), 6-hydroxykaempferol-3,6-di-*O*-β-D-glucopyranoside (**3**), 4-*O*-β-D-glucosyl-*trans-p*-coumaric acid (**5**), and 4-*O*-β-D-glucosyl-*cis-p*-coumaric acid (**6**). Their structures were identified on the basis of chemical and spectroscopic methods.

Keywords: *Carthamus tinctorius*; flavonoids; aromatic glucosides; chemical and spectroscopic methods

1. Introduction

The dried flower of safflower, *Carthamus tinctorius* L. is a traditional Chinese medicine widely used in china, having the function of promoting blood circulation by removing blood stasis.¹ The chemical constituents from this plant have been examined, and the isolation of flavonoids,^{2,3} polyacetylenes,⁴ serotonin derivatives,⁵ steroids,⁶ lignans,^{7,8} alkane diol,^{9,10} and colouring matter¹¹ have been reported. During the course of our phytochemical investigation, a new flavonoid and a new aromatic glucoside, together with four known compounds (Figure 1), were isolated from *C. tinctorius*. This paper describes the isolation and structural characterization of these compounds.

2. Results and discussion

Compound **1** was obtained as a yellow gummy material. The molecular formula was determined to be C₂₇H₃₂O₁₆ by the HR-FABMS

(*m/z* 613.1769 [M + H]⁺). The IR spectrum of **1** suggested the presence of conjugated carboxyl (1650 cm⁻¹) and hydroxyl groups, which were due to sugar moieties (3363 and 1075 cm⁻¹). The ¹H NMR spectrum of **1** showed the presence of the flavanone structure with a *p*-hydroxylphenyl group at δ 7.33 (2H, d, *J* = 8.4 Hz, H-2' and H-6') and 6.80 (2H, d, *J* = 8.4 Hz, H-3' and H-5'). The down-field chemical shift of OH-5 (δ 11.95) suggested that it should have formed an internal hydrogen bond with the carbonyl carbon C-4 (δ 197.7). Furthermore, the characteristic signals of flavanone were revealed at δ 5.48 (1H, dd, *J* = 13.1, 2.4 Hz) for H-2α, at δ 2.70 (1H, dd, *J* = 17.2, 2.4 Hz) for H-3α, and at 3.30 (1H, dd, *J* = 17.2, 13.1 Hz) for H-3β. Particularly, in the UV spectrum, the absorption band II at 280 nm and band I at 340 nm were observed, which were strong evidence for flavanones. Two anomeric protons at δ 4.71 (1H, d, *J* = 6.9 Hz, H-1'') and δ 4.92 (1H, d, *J* = 6.9 Hz, H-1''') demonstrated the

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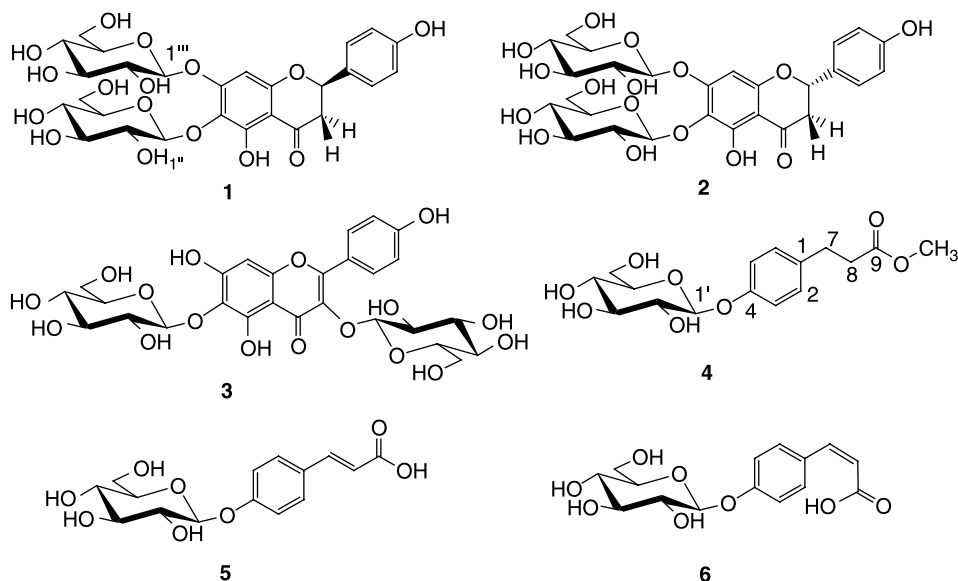


Figure 1. The structure of compounds **1**–**6**.

presence of sugar moieties. Acid hydrolysis indicated the existence of glucose. The coupling constant ($J = 6.9\text{ Hz}$) of the anomeric protons indicated that the two glucosyl moieties exhibited a β -configuration. The above facts suggested that **1** should be a glucoside of flavanone. In the HMBC experiment of **1** (Figure 2), the correlations of H-1'' and C-6 as well as H-1''' and C-7 confirmed that two glucosyl moieties were

attached at C-6 and C-7, respectively. The absolute configuration at C-2 was determined as *R* by observing the CD spectrum, which displayed a negative Cotton effect at 345 nm and a positive Cotton effect at 290 nm.¹² On the basis of the above evidence, the structure of **1** was elucidated to be (2*R*)-4',5-dihydroxy-6,7-di-*O*- β -D-glucopyranosyl flavanone.

Compound **2** was isolated as a white amorphous powder. The ^1H NMR and ^{13}C

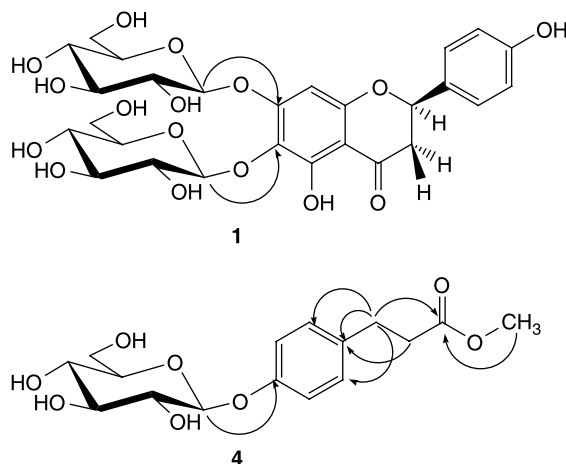


Figure 2. The key HMBC correlations of compounds **1** and **4**.

NMR data were very similar to those of **1**. But the absolute configuration at C-2 was determined as *S* by observing the CD spectrum, which displayed a positive Cotton effect at 345 nm and a negative Cotton effect at 290 nm. Furthermore, by comparison of their spectral data (^1H NMR and ^{13}C NMR) with those reported in the literature,¹² the structure of **2** was elucidated to be (2*S*)-4',5-dihydroxyl-6,7-di-*O*- β -D-glucopyranosyl flavanone.

Compound **4** was isolated as a white amorphous powder. The molecular formula was determined to be $\text{C}_{16}\text{H}_{22}\text{O}_8$ by HR-FABMS (m/z 343.1393 [$\text{M} + \text{H}$]⁺). The IR spectrum of **4** showed absorption bands at 3426, 1732, 1560, and 1458 cm^{-1} , suggesting the presence of hydroxyl group, ester carbonyl group, and aromatic rings. The ^1H NMR spectrum of **4** showed signals assignable to a *p*-substituted benzene ring at δ 6.93 (2H, d, $J = 8.4$ Hz) and 7.13 (2H, d, $J = 8.4$ Hz), two methylenes at δ 2.59 (2H, t, $J = 7.8$ Hz, H-8), 2.79 (2H, t, $J = 7.8$ Hz, H-7), indicating the existence of a $-\text{CH}_2-\text{CH}_2-$ unit, and a methoxyl at δ 3.57 (3H, s). Acid hydrolysis indicated the existence of glucose, which was confirmed from the anomeric proton at δ 4.80 (1H, d, $J = 7.5$ Hz, H-1') and the corresponding carbon signal for the anomeric carbon at δ 100.6 (C-1'). The coupling constant ($J = 7.5$ Hz) of the glucose anomeric proton indicated that the anomeric configuration was β -oriented. In the HMBC spectrum (Figure 2), the correlations of H-1', H-6, and H-2 with C-4 suggested that the glucosyl was linked to C-4. The correlations of H-7 with C-1, C-2, and C-6, and H-8 with C-1 indicated that the $-\text{CH}_2-\text{CH}_2-$ unit was linked to C-1. The correlations of H-7 and H-8 with the carbonyl carbon indicated that the carbonyl carbon (C-9) was attached to C-8 (δ 35.3). The correlation of methoxyl protons with the carbonyl carbon suggested that the $\text{CH}_3\text{O}-$ was attached to C-9. On the basis of the above evidence, the structure of **4** was elucidated to be methyl-3-(4-*O*- β -D-glucopyranosylphenyl) propionate.

6-Hydroxykaempferol-3,6-di-*O*- β -D-glucopyranoside (**3**), 4-*O*- β -D-glucosyl-*trans-p*-coumaric acid (**5**), and 4-*O*- β -D-glucosyl-*cis-p*-coumaric acid (**6**) were identified by comparison of their spectral data (^1H NMR and ^{13}C NMR) with those reported in the literature.¹³⁻¹⁵

3. Experimental

3.1 General experimental procedures

Optical rotations were obtained on a Perkin-Elmer 241MC polarimeter. IR spectra were taken on a Bruker IFS-55 infrared spectrophotometer. UV spectra were obtained on a Shimadzu UV-260 spectrophotometer. The NMR data were recorded on Bruker AV-600 (600 MHz for ^1H and 150 MHz for ^{13}C) in $\text{DMSO}-d_6$ with TMS as the internal standard. The HR-FABMS data were obtained using the Micross Mass Autospec-UltimaE TOF mass spectrophotometer. Chromatography was performed on silica gel (200–300 mesh, Qingdao Haiyang Chemical Factory, Qingdao, China), Sephadex LH-20 (Pharmacia), reversed-phase HPLC (Shimadzu LC-10A vp).

3.2 Plant material

Dried petals of *C. tinctorius*, cultivated in the Xinjiang Province of China, were bought from the Corporation of Traditional Chinese Medicine of Shenyang, China, in June 2005. A voucher specimen was identified by Professor Qi-shi Sun and has been deposited at the School of Traditional Chinese Medicine of Shenyang Pharmaceutical University, China (No. 6025).

3.3 Extraction and isolation

Dried petals (5 kg) of *C. tinctorius* were extracted three times with hot 95% EtOH, every time for 2 h, and the combined solution was concentrated *in vacuo* to a syrup (1200 g), followed by suspension in water. The suspension was extracted with petroleum ether, ethyl acetate, and *n*-butanol successively.

Table 1. ^{13}C NMR spectral data of compounds **1**, **2** and **4** in $\text{DMSO}-d_6$ (150 MHz).

Position	1	2	4
1			133.8
2	78.9	78.9	129.2
3	42.4	42.4	116.2
4	197.7	197.7	160.0
5	154.6	154.8	116.2
6	127.1	127.0	129.2
7	158.1	158.2	29.5
8	94.9	94.9	35.2
9	158.5	158.6	172.8
9-OCH ₃			51.4
10	103.5	103.5	
1'	128.6	128.8	100.6
2'	128.6	128.6	73.3
3'	115.3	115.3	77.1
4'	157.9	157.9	69.8
5'	115.3	115.3	76.7
6'	128.6	128.6	60.8
1''	103.8	103.9	
2''	73.4	73.4	
3''	76.4	76.4	
4''	69.7	69.7	
5''	77.2	77.3	
6''	60.7	60.7	
1'''	100.7	100.7	
2'''	74.2	74.2	
3'''	76.4	76.4	
4'''	69.8	69.9	
5'''	77.2	77.3	
6'''	60.9	60.9	

The *n*-butanol fraction (200 g) was further chromatographed over a D101 macroporous resin column eluted with H_2O , 30, 70, and 95% EtOH gradually. The fraction (60 g) eluted with 30% EtOH was subjected to silica gel column chromatography (eluted with CHCl_3 and MeOH in increasing polarity) to obtain nine fractions (I–IX). Fraction IV was purified using Sephadex LH-20 column chromatography and preparative HPLC to obtain compounds **1** (10 mg), **2** (25 mg), **3** (50 mg), **4** (12 mg), **5** (29 mg), and **6** (18 mg).

3.3.1 (2R)-4',5-Dihydroxyl-6,7-di-O- β -D-glucopyranosyl flavanone (**1**)

Yellow gummy material, $[\alpha]_D^{25} - 21.6$ (MeOH). IR (KBr) ν_{max} (cm^{-1}) 1650, 3363,

1075; UV (nm) 280, 340; ^1H NMR ($\text{DMSO}-d_6$) δ : 7.33 (2H, d, $J = 8.4$ Hz, H-2' and H-6'), 6.80 (2H, d, $J = 8.4$ Hz, H-3' and H-5'), 6.37 (1H, s, H-8), 11.95 (OH-5), 5.48 (1H, dd, $J = 13.1, 2.4$ Hz, H-2 α), 2.70 (1H, dd, $J = 17.2, 2.4$ Hz, H-3 α), 3.30 (1H, dd, $J = 17.2, 13.1$ Hz, H-3 β). 4.71 (1H, d, $J = 6.9$ Hz, H-1''), 4.92 (1H, d, $J = 6.9$ Hz, H-1'''); ^{13}C NMR spectral data, see Table 1; HR-FABMS m/z : 613.1780 [$\text{M} + \text{H}$] $^+$ (calcd for $\text{C}_{27}\text{H}_{33}\text{O}_{16}$, 613.1769).

3.3.2 Methyl-3-(4-O- β -D-glucopyranosylphenyl) propionate (**4**)

White amorphous powder, $[\alpha]_D^{25} - 25.8$ (MeOH). IR (KBr) ν_{max} (cm^{-1}) 1585, 1490; ^1H NMR ($\text{DMSO}-d_6$) δ : 7.13 (2H, d, $J = 8.4$ Hz, H-2 and H-6), 6.93 (2H, d, $J = 8.4$ Hz, H-3 and H-5), 2.59 (2H, t, $J = 7.8$ Hz, H-8), 2.79 (2H, t, $J = 7.8$ Hz, H-7), 3.57 (3H, s, 9-OCH₃), 4.80 (1H, d, $J = 7.5$ Hz, H-1'); ^{13}C NMR spectral data, see Table 1; HR-FABMS m/z : 343.1405 [$\text{M} + \text{H}$] $^+$ (calcd for $\text{C}_{16}\text{H}_{23}\text{O}_8$, 343.1393).

3.4 Acid hydrolysis of **1** and **4**

Compound **1** (5 mg) was refluxed with 2 N HCl in aqueous MeOH (5 ml) for 8 h. The reaction mixture was diluted with water (10 ml) and extracted with EtOAc. The aqueous part was neutralized with a saturated solution of Na_2CO_3 and filtered. The filtrate was concentrated under reduced pressure and examined for sugar identification on PC with an authentic sample of glucose. The methods of acid hydrolysis of **4** were the same as that of **1**.

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